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THE VALUE OF ΔG° FOR THE HYDROLYSIS OF ATP*

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SUMMARY

I. The equilibrium constant of the reaction catalysed by glutamine synthetase (L-glutamate: ammonia ligase (ADP), EC. 6.3.1.2),

$$K_{\rm obs} = \frac{[\rm Gln] \cdot [\rm ADP] \cdot [\rm P_i]}{[\rm Glu] \cdot [\rm ammonia] \cdot [\rm ATP]} \; ,$$

was measured at 37 °C between pH 6.6 and 7.6 and [Mg²⁺] between I and 34 mM.

- 2. The stability constant of the magnesium–glutamate complex was found to be 7.66 \pm 0.26 (S.E.) M⁻¹ at I 0.2 and 37 °C.
- 3. By combining these results with published data for the equilibrium of the glutaminase reaction, the pK and binding constants of the magnesium complexes with P_i , ADP and ATP, activity coefficients and enthalpy data, the $-\Delta G^{\circ}$ of the reaction

$$ATPH^{3-} + H_2O \rightleftharpoons ADPH^{2-} + H_2PO_4^-,$$

at I o and 25 °C was calculated to be 7.53 \pm 0.09 (S.E.) kcal/mole (31.5 \pm 0.4 kJ/mole).

- 4. This value is 2.33 kcal/mole less than that calculated by R. C. Phillips, P. George and R. J. Rutman ($J.\ Biol.\ Chem.$, 244 (1969) 3330). The difference is caused mainly by (i) the lower value used by us for the equilibrium constant of the glutamine synthetase reaction, (ii) a lower value used for the stability constant of the magnesium-glutamate complex, (iii) allowing for the effect of lowering the temperature from 37 to 25 °C on the RT term.
- 5. Calculations from data in the literature on the equilibria of the reactions catalysed by hexokinase and glucose-6-phosphatase yield a value similar to that reported above.
- 6. Data are tabulated and plotted graphically for the value of $-\Delta G^{\circ}_{obs}$ at 25 and 37 °C, between pH 6 and 9 and for Mg²+ concentrations up to 50 mM. At pH 7.5, I 0.2 and 25 °C, ΔG°_{obs} is -8.48 kcal/mole (-35 kJ/mole), 2.26 kcal/mole less negative than that calculated by Phillips *et al*.

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^{*} Dedicated to Professor H. Veldstra on the occasion of his retirement from the chair of biochemistry of the University of Leiden.

INTRODUCTION

Since the energy required for a large number of biochemical processes is provided by the hydrolysis of ATP to ADP and P_i , the value for the change in the standard free energy, ΔG° , of the reaction

$$ATP + H_2O \rightleftharpoons ADP + P_i \tag{1}$$

is of fundamental importance. The only method available to determine this quantity is from measurements of the equilibrium constant

$$K_{\text{obs}} = \frac{[\text{ADP}]_{\text{e}} \cdot [\text{P}_{\text{i}}]_{\text{e}}}{[\text{ATP}]_{\text{e}}},$$

in which the symbols $[ADP]_e$, $[P_i]_e$ and $[ATP]_e$ refer to the total concentrations of the compounds, made up of the various ionic species, including those complexed with magnesium, present at equilibrium. The corresponding change of standard free energy, ΔG°_{obs} , is equal to $-RT \ln K_{obs}$. Since the ionisation constants and the stability constants of the magnesium complexes differ for products and reactants, ΔG°_{obs} is a function of pH and Mg^{2+} concentration.

The value of $K_{\rm obs}$ is so high that equilibrium of Eqn I is reached at vanishingly small concentrations of ATP, thus precluding the direct measurement of the equilibrium constant. It is possible, however, to calculate it indirectly by combining data for an ATP-driven synthesis and the hydrolysis of the synthesized product. For example, the sum of Eqns 2 and 3

Glutamate + ammonia + ATP
$$\rightleftharpoons$$
 glutamine + ADP + P_i (2)

Glutamine +
$$H_2O \rightleftharpoons glutamate + ammonia$$
 (3)

is Eqn 1, so that $K_{obs} = K_a \cdot K_b$ where K_a and K_b are the equilibrium constants of the reactions shown in Eqns 2 and 3, respectively.

By combining the data of Levintow and Meister¹ for glutamine synthesis and of Benzinger $et~al.^2$ for glutamine hydrolysis, together with new values for ionisation constants and stability constants for magnesium complexes, Alberty³,⁴ and Phillips $et~al.^5$ have calculated $\Delta G^{\circ}_{\rm obs}$ at various temperatures, Mg²+ concentrations, ionic strength and pH values.

The accuracy of these calculations depends upon the reliability of the primary equilibrium data. In view of the fundamental importance of having an accurate value for ΔG°_{obs} , for example for the determination of the phosphate potential of oxidative or photosynthetic phosphorylation^{6–11}, we thought it desirable to reexamine the value for the equilibrium of the glutamine synthetase reaction used by Alberty^{3, 4} and Phillips *et al.*⁵ in their calculations. Since a value differing considerably from that reported by Levintow and Meister¹ was obtained, a reassessment of the value of ΔG°_{obs} was necessary.

The value for $-\Delta G^{\circ}_{obs}$, calculated from our data, including also a new determination of the stability constant of the magnesium–glutamate complex near pH 7, is, at pH 7.5, I 0.2 and 25 °C and low Mg²⁺ concentration, 2.3 kcal/mole less negative than that calculated by Phillips *et al.*⁵.

RESULTS

The equilibrium constant of the glutamine synthetase reaction

The measurements of the glutamine synthetase reaction were carried out at 37 °C and at various values of pH, ionic strength and Mg²⁺ concentration. In order to reduce the time required to reach equilibrium, all reactants and products were present at zero time and the increase or decrease of the concentration term

$$\frac{[Gln] \cdot [ADP] \cdot [P_i]}{[Glu] \cdot [ammonia] \cdot [ATP]}$$

was followed until a constant value was reached. Illustrative experiments, carried out at pH 6.6, I 0.2 and 50 mM magnesium, are shown in Table I and Fig. I. In three different experiments, decreasing initial values of the concentration term were used. Within experimental error, the same value was reached after 60 min. The 6 values obtained by combining the data at 60 min and 120 min, together with 6 additional values not shown in Fig. I, give a mean value for $K_{\rm a~obs}$ of 162 \pm 5 (S.E.). Fig. 2 shows the results of an experiment, carried out at pH 7.0, I 0.3 and 50 mM magnesium, in which the equilibrium was approached from both sides.

The results of these and other experiments under different conditions are summarized in Table II. The results differ considerably from those reported by Levintow and Meister¹ (1200 at pH 7.0) and Varner and Webster¹² (1700 at pH 7.4).

The stability constant of the magnesium-glutamate complex

The value of the stability constant of the magnesium-glutamate complex used by Alberty^{3,4} and Phillips *et al.*⁵ is that reported by Lumb and Martell¹³. However, this value refers to the complex with zero charge, whereas at pH values between

TABLE I
DETERMINATION OF THE EQUILIBRIUM CONSTANT OF THE GLUTAMINE SYNTHETASE REACTION

The reaction mixture contained 100 mM Tris, 50.4 mM MgCl₂, 25 mM 2-mercaptoethanol and ATP, ADP, P_I, glutamate, glutamine and ammonia in the amounts given in the table for zero time. The solution was brought to pH 6.6 by addition of KOH. The ionic strength was 0.2. The reactions were carried out in a volume of 3 ml in centrifuge tubes placed in a Dubnoff shaker at 37 °C. The reaction was started by the addition of 1.6 units (μ mole/min) glutamine synthetase and stopped by the addition of 0.3 ml 40 % HClO₄ and 0.1 ml 5 % serum albumin. The protein was removed by centrifugation and the supernatant was neutralized with 0.1 M Tris–10 % (w/v) KOH, after which the KClO₄ was removed by freezing and thawing.

Expt	Reaction time (min)	[Glu] (mM)	[Ammonia] (mM)	[ATP] (mM)	[Gln] (mM)	[ADP] (mM)	$[P_1]$ (mM)	$ \begin{array}{c} [Gln] \cdot [ADP] \cdot [P_1] \\ \hline [Glu] \cdot [ammonia] \cdot [ATP] \end{array} $
1	0	1.00	0.97	0.97	8.8	9.0	9.1	774
	60	1.49	1.73	1.56	8.3	8.4	9.5	165
	120	1.66	1.70	1.62	8.5	8.2	9.5	145
2	o	1.33	1.36	1.36	8.8	9.1	10.0	327
	60	1.55	1.91	1.59	8.3	8.5	10.2	149
	120	1.73	1.90	1.75	8.8	9.5	10.2	147
3	0	1.66	1.68	1.62	8.8	8.8	10.2	175
-	60	1.60	1.73	1.61	8.6	9.2	9.7	171
	120	1.75	2.14	1.70	8.7	9.0	10.4	129

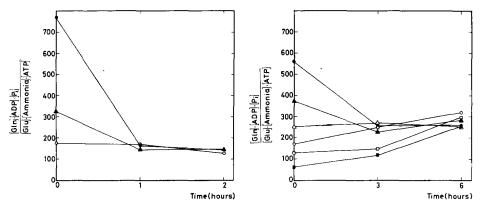


Fig. 1. Determination of the equilibrium constant of the glutamine synthetase reaction at 37 $^{\circ}$ C, pH 0.6, I 0.20 and 50.4 mM MgCl₂. Details are given in Table I.

Fig. 2. Determination of the equilibrium constant of the glutamine synthetase reaction at 37 $^{\circ}$ C, pH 7.0, I 0.30 and 50.4 mM MgCl₂. The experiment was carried out in the same way as in Table I.

TABLE II

DETERMINATION OF THE EQUILIBRIUM CONSTANT OF THE GLUTAMINE SYNTHETASE REACTION UNDER DIFFERENT CONDITIONS

The mean values reached at equilibrium, under the conditions specified, are given. Temperature, $37\,^{\circ}\text{C}$.

pΗ	I	$egin{array}{c} [K] \ (mM) \end{array}$	$[Mg] \ (mM)$	[ATP] (mM)	[ADP] (mM)	$[P_i]$ (mM)	$[Mg^{2+}] \ (mM)$	Ka* obs
6,6	0.20	45	50.4	1.6	8.8	10.2	34	162
7.0	0.30	98	50.4	1.3	8.8	9.7	34	270
7.1	0.22	93	10.7	1.3	7.6	10.1	2,2	280
7.5	0.21	84	49.5	0.9	9.3	9.4	33	668
7.58	0.17	73	9.8	0.8	9.2	1.01	1.3	831

^{*} $K_{a \text{ obs}} = ([Gln] \cdot [ADP] \cdot [P_i]) / ([Glu] \cdot [ammonia] \cdot [ATP])$

6.6 and 7.5, where the measurements of the equilibrium constant of the glutamine synthetase reaction have been carried out, the complex carries one positive charge:

$$Glu^{2-,1+} + Mg^{2+} \rightleftharpoons Mg - Glu^{+}$$

$$\tag{4}$$

The equilibrium constant of this reaction was determined in two ways, by titration and by an ion-exchange method. In the pH range 3-7, the equilibria given by Eqns 4-6 have to be taken into account.

$$Glu^+ \rightleftharpoons Glu^{-+} + H^+$$
 (5)

$$Glu^{-+} \rightleftharpoons Glu^{2-,1+} + H^{+}$$
 (6)

Putting

$$[Mg] = [Mg^{2+}] + [Mg-Glu^{+}]$$

 $[Glu] = [Glu^{+}] + [Glu^{-+}] + [Glu^{2-,1+}] + [Mg-Glu^{+}]$

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then

$$[Mg-Glu^{+}] = [Glu] - \frac{[Glu^{2-,1+}][(H^{+})^{2} + K_{Glu^{+}} \cdot (H^{+}) + K_{Glu^{+}} \cdot K_{Glu^{-+}}]}{K_{Glu^{+}} \cdot K_{Glu^{-+}}}$$
(7)

where K_{Glu}^+ and K_{Glu}^{-+} are the equilibrium constants of the reactions given in Eqns 5 and 6, respectively, and (H+) is the activity of H+.

At pH 7, practically all the glutamate is present as Glu^{2-,1+}. If we titrate a solution at this pH with HCl, the concentration of glutamate that becomes protonated, $\lceil Glu^{-+} \rceil + \lceil Glu^{+} \rceil$, will equal $[HCl]_{added}$ — (H^{+}) , i.e.

$$[Glu^{-+}] + [Glu^{+}] = [Glu^{2-,1+}] \left(\frac{(H^{+})}{K_{Glu^{-+}}} + \frac{(H^{+})^{2}}{K_{Glu^{-+}}} \right)$$

i.e.

$$[Glu^{2-,1+}] = \frac{K_{Glu^+} \cdot K_{Glu^-} + \{[HCl]_{added} - (H^+)\}}{K_{Glu^+} \cdot (H^+) + (H^+)^2}$$
(8)

If

$$\bar{n} = \frac{[\mathrm{Mg-Glu}^+]}{[\mathrm{Mg}]},$$

$$K_{f} = \frac{[Mg - Glu^{+}]}{[Mg^{2+}] \cdot [Glu^{2-,1+}]} = \frac{\bar{n}}{(1 - \bar{n}) \cdot [Glu^{2-,1+}]}$$
(9)

The pK values were determined by titration in the absence of magnesium and \bar{n} was calculated from Eqn 10

$$\bar{n} = \frac{[Glu] - [Glu^{2^{-,1^{+}}}] \{ (H^{+})^{2} + K_{Glu^{+}} \cdot (H^{+}) + K_{Glu^{+}} \cdot K_{Glu^{-+}} \} / K_{Glu^{+}} \cdot K_{Glu^{-+}}}{[Mg]}$$
(10)

and $K_{\mathbf{f}}$ from Eqn 9.

TABLE III DETERMINATION OF STABILITY CONSTANT OF Mg-GLUTAMATE+ BY TITRATION

50.9 ml of a solution containing 33.9 mM sodium hydrogen glutamate and 57 mM MgCl₂ was titrated with 0.506 M HCl. I 0.2. 25 °C. $K_{\rm Glu}^+=5.5\cdot 10^{-3}$ M. $K_{\rm Glu}^{-+}=7.07\cdot 10^{-5}$ M.

HCl $added$ (ml)	pΗ	$[Glu^{2-,1+}]^{\star}\ (mM)$	\overline{n} * *	K_f^{***} (M^{-1})
0	6.22			
0.2	5.20	22.I	0.168	9.15
0.4	4.90	22.0	0.134	7.03
0.6	4.70	20.8	0.119	6.52
0.8	4.50	17.3	0.145	9.76
1.0	4.40	17.1	0.113	7.43
1.2	4.29	15.9	0.099	6.96
1.6	4.08	12.9	0.082	6.98
1.8	3.98	11.3	0.075	7.11
2.0	3.88	9.94	0.065	7.03 Mean 7.55 ± 0.37 (S.E.)

^{*}Calculated by Eqn 8.

*Calculated by Eqn 10.

**Calculated by Eqn 9.

A typical titration at 25 °C and I=0.2 is given in Table III. The average values for this and two other ionic strengths are given in Table IV. The stability constant at I 0.2 for Mg–Glu⁺, 7.5 M⁻¹, is much less than the value of 80 M⁻¹ given for Mg–Glu° by Lumb and Martell¹³ and used by Alberty^{3,4} and Phillips *et al.*⁵.

The value obtained by the titration method was confirmed by the ion-exchange procedure of Schubert¹⁴. Using this method, $K_{\rm f}$ is given by the expression

$$\left(\frac{K_{\rm d}^{\circ}}{K_{\rm d}}-1\right)/[{\rm Glu}]^n$$

where K°_{d} and K_{d} are the distribution coefficients of Mg²⁺ between the ion exchanger and the solution in the absence and presence of glutamate, respectively, and n is the ratio of the number of molecules of metal to ligand in the complex. Assuming n = 1, a value of 7.66 \pm 0.26 (S.E.) M⁻¹ at 37 °C and I = 0.2 was obtained (Table V).

Table IV stability constant of Mg-glutamate⁺ at different ionic strengths, as determined by titration

I	$K_{ m G1u}^+ \ (M)$	$K_{\mathbf{G1u}}^{-+}$ (M)	K_f^* (M^{-1})
0.08 0.2 0.27	4.4·10 ⁻³ 5.5·10 ⁻³ 5.5·10	4.9·10 ⁻⁵ 7.1·10 ⁻⁵	25.4 ± 1.1 7.5 ± 0.4 8.1 ± 0.3

^{*} Mean \pm S.E. of mean.

Method as in Table III.

TABLE V

DETERMINATION OF THE STABILITY CONSTANT OF Mg-GLUTAMATE+ BY ION-EXCHANGE METHOD

100 mg Dowex 50-X 8–400 (Sigma) were suspended in 10 ml 15 mM sodium diethylbarbiturate buffer containing 50 μ M MgCl₂ and varying amounts of sodium hydrogen glutamate and NaCl to bring the ionic strength to 0.21. The pH was brought to 7.4 with HCl and the suspension shaken in a Dubnoff shaker for 3–5 h at 37 °C. Magnesium was then determined with a Perkin–Elmer Atomic Absorption Spectrophotometer Model 305. $K_{\rm d}^{\circ}$, the distribution coefficient of Mg²+between the ion exchanger and the solution in the absence of glutamate, determined by extrapolation of $K_{\rm d}$, the distribution coefficient in the presence of glutamate, to zero glutamate concentration, was equal to 0.23 ml·mg⁻¹.

$egin{array}{c} [Glu] \ (mM) \end{array}$	$[Mg^{2+}] \ (\mu M)$	$_{(ml\cdot mg^{-1})}^{K_d}$	$K_f^* (M^{-1})$
39	17.5	0.186	6.08
58	19.4	0.158	7.08
78	20.7	0.142	7.96
117	22.5	0.122	7.58
156	25.0	0.100	8.34
195	26.3	0.0905	7.91
39	18.8	0.171	8.42
78	20.0	0.150	6.85
117	22.5	0.122	7.58
156	25.0	0.100	8.34 Mean 7.66 ± 0.26 (S.E.)

^{*} $K_{\mathbf{f}} = (K_{\mathbf{d}}^{\circ}/K_{\mathbf{d}} - \mathbf{I})/[Glu].$

Calculation of ΔG°_{obs} for the hydrolysis of ATP

Since the ionisation constants and the stability constants of the magnesium complexes differ for products and reactants, ΔG°_{obs} will be a function of pH and $[Mg^{2+}]$. At the high concentrations of K⁺ used in our experiments, it is also necessary to consider binding of the reactants and products to this cation. In what follows, we shall restrict ourselves to the pH range 6–9, normally the physiological region. It will be assumed that the pK for the adenine ring is the same in both ATP and ADP. Alberty et al.¹⁵ found that this pK equals 4 in both compounds, and Phillips et al.¹⁶ found that Mg²⁺ has no effect.

Between pH 6 and q, the following equilibria for ATP have to be considered:

The values for the equilibrium constants at 37 $^{\circ}$ C, as given in or calculated from the literature, are shown in Table VI, which also includes data for the corresponding equilibrium constants for ADP and P_i .

From the equilibrium equations we may derive (cf. ref. 5)

$$y_{\text{ATP}} = \frac{[\text{H}^+]}{K_4[\text{Mg}^{2+}](\text{H}^+) + K_1 K_{\text{K}_1}[\text{K}^+] + K_1 K_2[\text{Mg}^{2+}] + (\text{H}^+) + K_1}$$

For phosphate the equilibria yield

$$y_{P_1} = \frac{(H^+)}{(H^+) + K_8 + K_9 K_{10} [Mg^{2+}] + K_9 K_3 [K^+]}$$

In agreement with Phillips et al.5 we shall use Eqn 11

$$ATPH^{3-} + H_2O \rightleftharpoons ADPH^{2-} + H_2PO_4^-$$
 (11)

as reference. For this equation

$$K_{\text{ref}} = \frac{[\text{ADPH}^{2-}] [\text{H}_2 \text{PO}_4^-]}{[\text{ATPH}^{3-}]}$$
 (12)

IONISATION AND STABILITY CONSTANTS FOR ATP, ADP AND P1 AT 25 °C AND 37 °C AND VARIOUS IONIC STRENGTHS TABLE VI

ATPH3- = ATP4- + H+	Constant	I o		I 0.17	I~o.2o	I 0.21	I 0.22	I 0.30	Keference
$ATPH^3- \leq ATP^4- + H^+$		25 °C	37 °C	37 °C	37 °C	37 °C	37 °C	3° 7°	
	K_1	2.09.10-8	1.86.10-8	1.12.10-7	1.10.10-7	1.10.10-7	1.12.10-7	9.76.10-8	20
$ m ATP^{4-} + Mg^{2+} \leftrightarrows ATPMg^{2-}$	K_2	6.76.105	$9.35\cdot 10^{5}$	4.46.104	4.90.104	5.11.104	$5.36 \cdot 10^{4}$	8.70.104	91
${ m ATPHMg^-} \leftrightarrows { m ATPMg^{2-}} + { m H^+}$	K_3	4.57.10-6	4.90.10-6	6.17.10-6	5.90 · 10 -6	$5.75 \cdot 10^{-6}$	5.50.10-6	4.57.10-6	91
$ m ATPH^{3-} + Mg^{2+} \leftrightarrows ATPHMg^{-}$	K_4	3.90.103	4.47.103	$6.45\cdot 10^2$	7.10.102	7.40.102	7.60 · 102	1.15.103	91
$ ext{ADPH}^2-\leftrightarrows ext{ADP}^3-\dotplus ext{H}^+$	K_5	$6.31 \cdot 10^{-8}$	5.75.10-8	$1.95 \cdot 10^{-7}$	1.78.10-7	1.78.10-7	1.78.10-7	1.51.10-7	20
${ m ADP^{3-} + Mg^{2+}} \leftrightarrows { m ADPMg^{-}}$	K_{6}	1.86.104	2.46.104	3.46.103	3.80.103	$3.98 \cdot 10^{3}$	4.07.103	$6.15 \cdot 10^{3}$	91
${ m ADPHMg^{\circ}} \leftrightarrows { m ADPMg^{-}} + { m H^{+}}$	K_7	4.18.10-6	4.80.10-6	$5.63 \cdot 10^{-6}$	5.63.10-6	5.50.10-6	5.50.10-6	5.14.10-6	91
$ ext{ADPH}^{2-} + ext{Mg}^{2+} \leftrightarrows ext{ADPHMg}^\circ$	K_8	$2.82\cdot 10^2$	$3.02\cdot 10^2$	$1.00\cdot 10^2$	$1.05 \cdot 10^2$	$1.07\cdot 10^2$	$_{\rm I.IO\cdot IO^2}$	$1.38\cdot 10^2$	91
$\mathrm{H_2PO_4^-} \leftrightarrows \mathrm{HPO_4^{2-}} + \mathrm{H^+}$	K_9	$6.61\cdot 10^{-8}$	7.09.10-8	1.90.10-7	$2.00\cdot 10^{-7}$	$2.05\cdot 10^{-7}$	2.10.10 ⁻⁷	2.18.10-7	20
$\mathrm{HPO_4^{2-}} + \mathrm{Mg^{2+}} \leftrightarrows \mathrm{HPO_4Mg^\circ}$	K_{10}	$5.12\cdot 10^2$	$6.31\cdot 10^2$	$1.13\cdot 10^2$	$1.05 \cdot 10^2$	1.07.102	$1.07\cdot 10^2$	$_{\rm I.OI\cdot IO^2}$	26, 27
$ ext{ATP}^{4-} + ext{K}^{+} \leftrightarrows ext{ATPK}^{3-}$	K_{K_1}			91	15	15	15	11	28*, 29*
${ m ADP^{3-} + K^{+} \leftrightarrows ADPK^{2-}}$	K_{K2}			7.6	7.6	7.6	9.7	9.2	28*, 29*
$\mathrm{HPO_4^{2-}} + \mathrm{K^+} \leftrightarrows \mathrm{HPO_4^{K^-}}$	$K_{\mathbf{K}3}$			4.6	4.6	4.6	4.6	4.6	28*

* Values at 37 °C are calculated using enthalpy data given by Smith and Alberty²⁸.

From the observed equilibrium constants of the glutamine synthetase and glutaminase reactions we may calculate

$$K_{\text{obs}} = \frac{[ADP] \cdot |P_i|}{[ATP]} \tag{13}$$

where [ADP] etc. indicate total concentrations of all ionic species and complexes of ADP etc. Combining Eqns 12 and 13 yields

$$K_{\text{ref}} = \frac{[\text{ADPH}^{2-}]}{[\text{ADP}]} \cdot \frac{[\text{H}_2 \text{PO}_4^-]}{[\text{P}_i]} \cdot \frac{[\text{ATP}]}{[\text{ATPH}^{3-}]} \cdot K_{\text{obs}}$$

$$= \frac{y_{\text{ADP}} \cdot y_{\text{P}_1}}{y_{\text{ATP}}} \cdot K_{\text{obs}}$$
(14)

From Eqn 14, ΔG°_{ref} may be calculated and, by reversal of the process described above, ΔG°_{obs} values at various values of pH and [Mg²⁺] may be calculated.

In order to allow for binding of glutamate to Mg^{2+} , under the conditions of measurement of the equilibrium constant of the glutamine synthetase reaction, it is necessary to calculate the concentration of Mg^{2+} unbound to ATP, ADP and P_i . This was calculated as in ref. 3, with the additional correction for the binding of K^+ to ATP, ADP and P_i . The values are listed in the second last column of Table II.

Making use of the stability constant of the magnesium-glutamate complex found at 37 $^{\circ}$ C (7.66), we can calculate

$$\begin{split} &\frac{[Gln] \cdot [ADP] \cdot [P_i]}{[Glu^{2^+, 1^-}] \cdot [ammonia] \cdot [ATP]} = \frac{[Gln] \cdot [ADP] \cdot [P_i]}{[Glu] \cdot [ammonia] \cdot [ATP]} \cdot (1 + 7.66[Mg^{2^+}]) \\ &= (1 + 7.66[Mg^{2^+}]) K_{a \text{ obs}} \\ &\frac{(Gln)}{(NH_4Glu)^2} \cdot \frac{[ADP] \cdot [P_i]}{|ATP]} = \frac{\gamma_{Gln}}{\gamma_{NH_4Glu}^2} (1 + 7.66[Mg^{2^+}]) K_{a \text{ obs}} \end{split}$$

where the parentheses indicate activities and γ refers to activity coefficient. The values for the activity coefficients, listed in Table VII, are calculated from the data of Benzinger *et al.*².

Even at the higher concentrations of Mg^{2+} used in the experiments summarized in Table VII the amount of magnesium–glutamate complex was a very small fraction of the $[Mg^{2+}]$, calculated from the concentrations of ATP, ADP and P_i and the stability constants of Mg^{2+} with these compounds.

For $K_{\rm b}$ (Eqn 3) at 37 °C we have used the value 242 M. This has been calculated from the value of 320 M given by Benzinger *et al.*² for 37 °C, with two correction factors. First, a value of $\gamma_{\rm NH4Glu}$ for pH 7 has been used instead of the value appropriate for pH 5 used by Benzinger *et al.*². This resulted in an increase of $K_{\rm b}$ to 341. Secondly, the factor to convert $K_{\rm b}$ from 25 to 37 °C was calculated from the heat of

TABLE VII calculation of ΔG°_{ref} .

	Expt No.				
	I	2	3	4	5
pН	6.6	7.0	7.1	7.5	7.58
Ī	0.20	0.30	0.22	0.21	0.17
VG1n	0.98	0.97	0.98	0.98	0.99
/NH4Glu	0.69	0.655	0.675	0.685	0.705
$[\mathrm{Mg}^{2+}]$ (M)	0.0341	0.0341	0.0022	0.0329	0.0013
$K_{\mathbf{a}}$ obs	162 2	70 2	280	568 8	31
$K_{ m obs} = 242 \cdot (\gamma_{ m Gln}/\gamma^2_{ m NH_4Glu}) \cdot (1+7.6)$ VADP	1.04·10 ⁵ 10.3·10 ⁻³	1.99·10 ⁵ 3.08·10 ⁻³			
ypi yatp $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pi}})/(y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$	$1.68 \cdot 10^5$	0.0857 0.34 · 10 ⁻³ 1.54 · 10 ⁵	5.81·10 ⁻³		0.0851 3.86·10 ⁻⁸ 1.95·10 ⁵
ypi yatp $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pi}}) / (y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$ $-\Delta G^{\circ}_{\text{ref}}$, 37 °C = 1.419 log K_{ref} (kca	1.32·10 ⁻³ 1.68·10 ⁵ al/mole)	0.34·10 ⁻³	5.81·10 ⁻³ 1.89·10 ⁵	0.17·10 ⁻³ 1.02·10 ⁵	3.86°10 ⁻⁸ 1.95°10 ⁵
ypi yatp $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pi}}) / (y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$ $- \Delta G^{\circ}_{\text{ref}}$, 37 °C = 1.419 log K_{ref} (kc)	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41	0.34·10 ⁻⁸ 1.54·10 ⁵ 7.36	5.81·10 ⁻³	0.17.10-3	3.86.10-8
ypi yatp $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pi}}) / (y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41	0.34·10 ⁻⁸ 1.54·10 ⁵ 7.36	5.81·10 ⁻³ 1.89·10 ⁵	0.17·10 ⁻³ 1.02·10 ⁵	3.86°10 ⁻⁸ 1.95°10 ⁵
VPI VATP $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pi}})/(y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$ $-\Delta G^{\circ}_{\text{ref}, 37 ^{\circ}\text{C}} = 1.419 \log K_{\text{ref}} (\text{kcc}$ $-\Delta G^{\circ}_{\text{ref}, 25 ^{\circ}\text{C}} = 1.364 \log K_{\text{ref}} + 6$	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41	0.34·10 ⁻⁸ 1.54·10 ⁵ 7.36	5.81·10 ⁻³ 1.89·10 ⁵	0.17·10 ⁻³ 1.02·10 ⁵	3.86°10 ⁻⁸ 1.95°10 ⁵
VPI VATP $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pl}})/(y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$ $-\Delta G^{\circ}_{\text{ref}}$, 37 °C = 1.419 log K_{ref} (kcc $-\Delta G^{\circ}_{\text{ref}}$, 25 °C = 1.364 log K_{ref} + C	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41 0.23 (kcal/mo 7.36 2.35	0.34·10 ⁻³ 1.54·10 ⁵ 7.36 dle) 7.31 1.74	7.49 7.43 2.24	0.17·10 ⁻³ 1.02·10 ⁵ 7.10 7.06 2.29	3.86·10 ⁻⁸ 1.95·10 ⁵ 7.51 7.44 2.49
VPI VATP $K_{\text{ref}} = (y_{\text{ADP}} \cdot y_{\text{Pl}})/(y_{\text{ATP}}) \cdot K_{\text{obs}} (M)$ $-\Delta G^{\circ}_{\text{ref}}$, 37 °C = 1.419 log K_{ref} (kcc $-\Delta G^{\circ}_{\text{ref}}$, 25 °C = 1.364 log K_{ref} + C $V_{\text{ADPH}}^{2-}/V_{\text{ATPH}}^{3-}$ V_{H2PO4}^{-}	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41 0.23 (kcal/mo 7.36 2.35 0.66	0.34·10 ⁻³ 1.54·10 ⁵ 7.36 dle) 7.31 1.74 0.61	7.49 7.43 2.24 0.65	0.17·10 ⁻³ 1.02·10 ⁵ 7.10 7.06 2.29 0.655	3.86·10 ⁻⁸ 1.95·10 ⁵ 7.51 7.44
$egin{aligned} & ext{VPI} \ ext{VATP} \ ext{K}_{ ext{ref}} &= (y_{ ext{ADP}} \cdot y_{ ext{Pi}})/(y_{ ext{ATP}}) \cdot K_{ ext{obs}} \left(ext{M} ight) \ - egin{aligned} & ext{$-G$} \end{array} & \overset{\circ}{ ext{c}} & = 1.419 \log K_{ ext{ref}} \left(ext{kcc} ight) \end{aligned}$	1.32·10 ⁻³ 1.68·10 ⁵ al/mole) 7.41 0.23 (kcal/mo 7.36 2.35 0.66	0.34·10 ⁻³ 1.54·10 ⁵ 7.36 dle) 7.31 1.74 0.61	7.49 7.43 2.24 0.65	0.17·10 ⁻³ 1.02·10 ⁵ 7.10 7.06 2.29 0.655	3.86·10 ⁻⁶ 1.95·10 ⁵ 7.51 7.44 2.49

the reaction reported by Kitzinger and Hems¹⁷. $K_{\rm obs}$ was calculated from the expression

$$K_{\text{obs}} = \frac{[\text{ADP}] \cdot [P_i]}{[\text{ATP}]} = \frac{(\text{Gln}) \cdot [\text{ADP}] \cdot [P_i]}{(\text{NH}_4 \text{Glu})^2 \cdot [\text{ATP}]} \cdot K_b$$
$$= \frac{\gamma_{\text{Gln}}}{\gamma_{\text{NH}_4 \text{Glu}}^2} (1 + 7.66[\text{Mg}^{2+}]) K_{\text{a obs}} \cdot 242$$

From this value of K_{obs} , K_{ref} was calculated by use of Eqn 14, and $\Delta G^{\circ}_{\text{ref}}$ by the usual relation between ΔG° and K. These values refer to 37 °C, the temperature at which the measurements of the equilibrium of the glutamine synthetase reaction were carried out.

The $K_{\rm ref}$ for 25 °C was calculated by making use of the van 't Hoff isochore and the values for ΔH for the hydrolysis of ATP given by Kitzinger and Benzinger¹⁸ (—4.8 kcal/mole) and Podolsky and Morales¹⁹ (—4.7 kcal/mole), determined at pH 8. The $\Delta H_{\rm ref}$ was calculated from this value according to the equation

$$\Delta H_{\text{ref}} = -4.75 + \frac{K_1/(\text{H}^+)}{1 + K_1/(\text{H}^+)} \cdot \Delta H_1 - \frac{K_5/(\text{H}^+)}{1 + K_5/(\text{H}^+)} \cdot \Delta H_5 - \frac{K_9/(H^+)}{1 + K_9/(H^+)} \cdot \Delta H_9$$

where ΔH_1 , ΔH_5 and ΔH_9 (kcal/mole) are the changes of enthalpy corresponding to the ionization of ATPH³⁻, ADPH²⁻ and H₂PO₄⁻, respectively. These values were

calculated from the data of Phillips *et al.*²⁰. This yields $\Delta H_{\rm ref} = -5.9$ kcal/mole at I 0.2. In the absence of sufficient data, no correction was made for a possible effect of ionic strength or binding of K⁺, under the conditions of the enthalpy measurements^{18,19} (cf. ref. 5).

$$\log \frac{K_{\text{ref, 25 °C}}}{K_{\text{ref, 37 °C}}} = \frac{-\Delta H}{2.303 \cdot R} \cdot \frac{310 - 298}{310 \cdot 298}$$

$$= \frac{5.9 \cdot 10^3 \cdot 12}{2.303 \cdot 1.988 \cdot 310 \cdot 298}$$

$$-\Delta G_{\text{ref, 25 °C}}^{\circ} = 2.303 \cdot 1.988 \cdot 298 \cdot 10^{-3} \log K_{\text{ref, 25 °C}} \text{ kcal/mole}$$

$$= 2.303 \cdot 1.988 \cdot 298 \cdot 10^{-3} \log K_{\text{ref, 37 °C}} + \frac{5.9 \cdot 12}{310} \text{ kcal/mole}$$

$$= 2.303 \cdot 1.988 \cdot 298 \cdot 10^{-3} \log K_{\text{ref, 37 °C}} + 0.23 \text{ kcal/mole}$$

$$\therefore -\Delta G_{\text{ref}, 25^{\circ}\text{C}}^{\circ} + \Delta G_{\text{ref}, 37^{\circ}\text{C}}^{\circ} = -2.303 \cdot 1.988 \cdot 12 \cdot 10^{-3} \log K_{\text{ref}, 37^{\circ}\text{C}} + 0.23 \text{ kcal/mole}$$

This correction is treated in detail because it differs from that given by Phillips et al.⁵, who assumed that $-\Delta G^{\circ}_{\rm ref,\ 25} \circ_{\rm C} + \Delta G^{\circ}_{\rm ref,\ 37} \circ_{\rm C} = 0.19$ kcal/mole. This is the term that is a measure of the effect of temperature on $K_{\rm ref}$ (Phillips used the value of -4.75 kcal/mole for $\Delta H_{\rm ref}$) which, in accordance with the principle of Le Chatelier, increases on lowering the temperature of an exothermic reaction. This increase of K, which by itself causes the increase of $-\Delta G^{\circ}$ by 0.19 kcal/mole, is, however, more than counterbalanced by the decrease in RT on lowering the temperature.

The values given in Table VII are applicable for the ionic strengths given. To calculate to zero ionic strength, the values of K_{ref} given have to be multiplied by

which may be equated with $\gamma_{\text{ADPH}^{2-}} \cdot \gamma_{\text{H}_2\text{PO}_4} / \gamma_{\text{ADP}^{3-}}$, since it may be assumed that $\gamma_{\text{ATPH}^{3-}} = \gamma_{\text{ADP}^{3-}}$. The quotient $\gamma_{\text{ADPH}^{2-}} / \gamma_{\text{ADP}^{3-}}$ may be obtained from the relationship between pK_b and ionic strength determined by Phillips *et al.*²⁰. Values for $\gamma_{\text{H}_2\text{PO}_4}^{-}$ used in Table VII are the means of the values for KH_2PO_4 and NaH_2PO_4 given by Stokes²¹ and Scatchard and Breckenridge²². The mean value of the five determinations for $-\Delta G^\circ_{\text{ref}, 25} \circ_{\text{C}}$ at zero ionic strength is 7.53 kcal/mole with a standard error of 0.09. This value is 2.33 kcal/mole lower than that calculated by Phillips *et al.*⁵. Our value for $-\Delta G^\circ_{\text{ref}, 37} \circ_{\text{C}}$ is 2.08 kcal/mole lower than that given by Phillips *et al.*⁵.

DISCUSSION

The difference in 2.33 kcal/mole for $\Delta G^{\circ}_{ref, 25} \circ_{\mathbb{C}}$ between our value and that of Phillips *et al.*⁵ is caused by the following factors:

(1) The equilibrium constant for the glutamine synthetase reaction is lower by a factor of 5 than that determined by Levintow and Meister¹ and used by Phillips et al.⁵. This accounts for a difference of 1.0 kcal/mole at 37 °C.

- (2) The stability constant for the complex formed between Mg²⁺ and Glu^{2-,1+}, the species of glutamate present at the pH values used, is considerably smaller than that between Mg²⁺ and Glu²⁻ used by Phillips *et al.*⁵. At the high Mg²⁺ concentration used by Levintow and Meister¹, this accounts for 0.9 kcal/mole.
- (3) In converting the data from 37 °C to 25 °C, Phillips *et al.*⁵ did not allow for the effect of lowering temperature on the term RT in calculating ΔG° from K. This amounts to about 0.25 kcal/mole.
- (4) We introduced into our calculations binding of K+ to ATP, ADP and P_i, activity coefficients for glutamine, and the activity coefficient for ammonium glutamate appropriate for pH 7, in calculating the equilibrium of the glutaminase reaction. These points are, however, of minor importance.

Since a major contribution to the difference of 2.33 kcal/mole is the disagreement between our measurements and those of Levintow and Meister¹ of the equilibrium constant of the glutamine synthetase reaction, it is desirable to have independent data from which ΔG for the hydrolysis of ATP can be calculated. Data in the literature for the equilibrium of the hexokinase²³ and glucose-6-phosphatase²⁴ reactions lend themselves to such a calculation, if certain assumptions may be made.

- (1) The second ionisation constant of glucose 6-phosphate and the stability constant of the complex with magnesium are the same as the values given for glucose 1-phosphate, viz. 3.1·10⁻⁷ M (ref. 25), and 3.32·10² M⁻¹ (ref. 26) at zero ionic strength and 30 °C.
- (2) The change in the second ionisation constant with ionic strength is the same as that for the second ionisation constant of AMP²⁰.
- (3) The change with ionic strength of the stability constant of the magnesium complex with glucose 6-phosphate is the same as that with ADPH²⁻ (ref. 16).

Robbins and Boyer²³ found a value of

$$K'_{\text{obs}} = \frac{[\text{Glc-6-}P] [\text{ADP}]}{(\text{Glc}) [\text{ATP}]}$$

of 184 at pH 6.04, I 0.30 and 30 °C, with $[Mg^{2+}] = 80$ mM. Using Eqn 15

$$Glc + ATPH^{3-} \rightleftharpoons Glc - 6 - P^{-} + ADPH^{2-}$$
(15)

as the reference reaction, in the absence of Mg2+

$$K'_{\text{ref}} = K'_{\text{obs}} \cdot \frac{y_{\text{Glc-6-P}} \cdot y_{\text{ADP}}}{y_{\text{ATP}}}$$

This gives $K_{\rm ref}$ of 248 at I 0.3 and 30 °C and in the absence of any thermochemical data it will be assumed that this value also applies to 25 °C.

Atkinson et al.24 found a

$$K''_{obs} = \frac{(Glc) [P_i]}{[Glc-6-P]}$$

of 320 M at pH 7.0, I 0.30 and 25 °C in the absence of Mg²⁺.

Table VIII values of $-\varDelta G^{\circ}{}_{obs}$ (kcal/mole) at 25 °C for various ionic strengths, pH and [Mg²+] concentrations

In parentheses are g	iven the $-\Delta G^{\circ}_{obs}$	values in kJ.
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pH	I	$[Mg^{2+}]\ (mM)$				
		0	1.0	10.0	25.0	50.0
6.00	0.00	7.59 (31.77)				
6.00	0.10	7.25 (30.33)	6.58 (27.53)	6.17 (25 82)	6.18 (25.85)	
6.00	0.15	7.30 (30.54)	6.64 (27.76)	6.23 (26.05)	6.23 (26.05)	6.30 (26.34)
6,00	0.20	7.37 (30.86)	6.69 (27.97)	6.29 (26.32)	6.30 (26.34)	6.38 (26.08)
6.50	0.00	7.71 (32.27)				
6.50	0.10	7.44 (31.12)	6.51 (27.25)	6.31 (26.39)	6.42 (26.87)	
6.50	0.15	7.49 (31.33)	6.57 (27.49)	6.36 (26.60)	6.46 (27.02)	6.62 (27.69)
6.50	0.20	7.56 (31.62)	6.62 (27.70)	6.43 (26.91)	6.54 (27.38)	6.71 (28.09)
7.00	0.00	8.01 (33.51)				
7.00	0.10	7.82 (32.72)	6.69 (28.00)	6.66 (27.88)	6.87 (28.74)	
7.00	0.15	7.87 (32.91)	6.75 (28.25)	6.71 (28.06)	6.90 (28.85)	7.13 (29.82
7.00	0.20	7.93 (33.17)	6.80 (28.46)	6.79 (28.40)	6.99 (29.25)	7.23 (30.26
7.50	0.00	8.55 (35.77)				
7.50	0.10	8.38 (35.05)	7.12 (29.80)	7.20 (30.12)	7.46 (31.20)	
7.50	0.15	8.42 (35.23)	7.18 (30.06)	7.24 (30.28)	7.48 (31.28)	7.74 (32.40
7.50	0.20	8.48 (35.48)	7.23 (30.27)	7.32 (30.04)	7.58 (31.70)	7.85 (32.85
8.00	0.00	9.24 (38.67)				
8.00	0.10	9.02 (37.74)	7.71 (32.24)	7.83 (32.74)	8.11 (33.91)	
8.00	0.15	9.06 (37.90)	7.77(32.51)	7.86 (32.90)	8.12 (33.99)	8.40 (35.16
8.00	0.20	9.12 (38.16)	7.82 (32.71)	7.95 (33.26)	8.22 (34.41)	8.51 (35.62
8.50	0.00	9.96 (41.67)				
8.50	0.10	9.69 (40.54)	8.35 (34.96)	8.49 (35.52)	8.78 (36.72)	
8.50	0.15	9.73 (40.70)	8.42 (35.22)	8.53 (35.68)	8.80 (36.82)	9.09 (38.03
8.50	0.20	9.79 (40.96)	8.47 (35.42)	8.61 (36.04)	8.90 (37.22)	9.19 (38.45
9.00	0.00	10.66 (44.59)				
y.00	0.10	10.37 (43.38)	9.03 (37.76)	9.17 (38.36)	9.46 (39.56)	
9.00	0.15	10.41 (43.54)	9.09 (38.03)	9.20 (38.50)	9.47 (39.63)	9.76 (40.85
9.00	0.20	10.47 (43.80)	9.14 (38.23)	9.29 (38.87)	9.58 (40.06)	9.87 (41.31

Using Eqn 16

$$Glc-6-P^- + H_2O \rightleftharpoons Glc + H_2PO_4^-$$
 (16)

as the reference reaction

$$K_{\text{ref}}'' = K_{\text{obs}}'' \cdot \frac{y_{P_i}}{y_{Glc-6-P}}$$

This gives a K''_{ref} of 674 M at 25 °C and I 0.3.

$$K_{\text{ref}} = K'_{\text{ref}} \cdot K''_{\text{ref}}$$
$$= 248 \cdot 674 \text{ M}$$
$$= 1.67 \cdot 10^5 \text{ M}$$

corresponding to $-\Delta G^{\circ}_{\rm ref,\ 25\ ^{\circ}C}$ of 7.11 kcal/mole at I 0.3. Converting to I 0, $-\Delta G^{\circ}_{\rm ref,\ 25\ ^{\circ}C}$ equals 7.14 kcal/mole, which is only just outside the range of values shown in Table VII, and far from the value of 9.9 kcal/mole calculated by Phillips et $al.^{5}$.

Reversal of the calculation carried out to obtain $K_{\rm ref}$ from $K_{\rm obs}$ yields $-\Delta G^{\circ}_{\rm obs}$ values referring to various conditions (Tables VIII and IX). In Fig. 3 is plotted the variation of $-\Delta G^{\circ}_{\rm obs}$ with pMg at different pH values.

Table IX values of $-\varDelta G^\circ{}_{\rm obs}$ (kcal/mole) at 37 °C for various ionic strengths, pH and Mg³+ concentrations

In parentheses are given the $-\Delta G^{\circ}_{obs}$ values in kJ.

pΗ	I	$[Mg^{2+}]\ (mM)$				
		0	1.0	10.0	25.0	50.0
6.00	0,00	7.69 (32.16)				
6.00	0.10	7.25 (30.34)	6,48 (27.10)	6.14 (25.70)	6.21 (25.98)	
6.00	0.15	7.27 (30.41)	6.48 (27.10)	6.14 (25.70)	6.21 (25.98)	6.34 (26.51
6.00	0.20	7.32 (30.65)	6.48 (27.10)	6.19 (25.88)	6.27 (26.22)	6.42 (26.85
6.50	0,00	7.81 (32.69)				
6.50	0.10	7.48 (31.30)	6.46 (27.02)	6.36 (26.63)	6.56 (27.44)	
6.50	0.15	7.51 (31.41)	6.47 (27.07)	6.37 (26.65)	6.55 (27.40)	6.78 (28.35
6.50	0.20	7.57 (31.67)	6.50 (27.18)	6.44 (26.95)	6.64 (27.80)	6.89 (28.82
7.00	0.00	8.13 (34.00)				
7.00	0.10	7.92 (33.15)	6.71 (28.09)	6.81 (28.51)	7.10 (29.70)	
7.00	0.15	7.95 (33.28)	6.74 (28.22)	6.82 (28.54)	7.09 (29.65)	7.37 (30.85)
7.00	0.20	8.03 (33.59)	6.79 (28.43)	6.92 (28.94)	7.20 (30.13)	7.50 (31.38)
7.50	0.00	8.70 (36.40)				
7.50	0.10	8.53 (35.71)	7.21 (30.18)	7.42 (31.03)	7.75 (32.41)	
7.50	0.15	8.57 (35.85)	7.26 (30.36)	7.43 (31.07)	7.73 (32.34)	8.04 (33.65)
7.50	0.20	8.65 (36.18)	7.32 (30.63)	7.53 (31.52)	7.86 (32.87)	8.18 (34.21)
8.00	0.00	9.44 (39.49)				
8.00	0.10	9.22 (38.56)	7.84 (32.81)	8.09 (33.85)	8.43 (35.29)	
8.00	0.15	9.25 (38.70)	7.89 (33.02)	8.10 (33.88)	8.42 (35.23)	8.74 (36.56)
8.00	0.20	9.33 (39.04)	7.96 (33.32)	8.21 (34.35)	8.55 (35.76)	8.88 (37.14)
8.50	0.00	10.20 (42.66)				
8.50	0.10	9.92 (41.50)	8.53 (35.67)	8.79 (36.76)	9.18 (38.40)	
8.50	0.15	9.95 (41.63)	8.58 (35.88)	8.79 (36.76)	9.14 (38.24)	9.46 (39.57)
8.50	0,20	10.03 (41.98)	8.65 (36.19)	8.91 (37.28)	9.27 (38.79)	9.60 (40.17
9.00	0.00	10.93 (45.73)				
9.00	0.10	10.63 (44.46)	9.23 (38.60)	9.49 (39.72)	9.85 (41.20)	
9.00	0.15	10.66 (44.59)	9.28 (38.82)	9.50 (39.75)	9.83 (41.14)	10.17 (42.54)
9.00	0.20	10.74 (44.94)	9.35 (39.13)	9.62 (40.23)	9.96 (41.67)	10.31 (43.14)

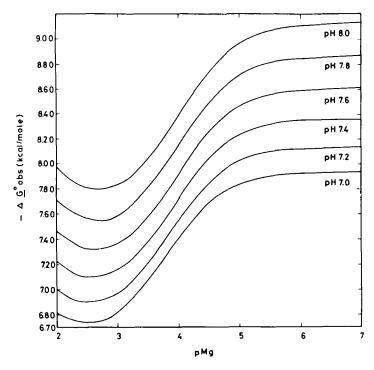


Fig. 3. ΔG°_{obs} as function of pMg at I 0.20, 25 °C and various pH values.

METHODS

Glutamine synthetase (L-glutamate: ammonia ligase (ADP), EC 6.3.1.2) was isolated from sheep brain by the method of Ronzio *et al.*³⁰ up to the step involving chromatography on DEAE-cellulose. This yields a fairly active preparation with an activity equivalent to 5 units (μ mole/min) per mg protein, determined by the method of Ronzio *et al.*³⁰. It was free from ATPase, adenylate kinase and glutaminase activity.

Phosphate was determined by the method of Sumner³¹.

ATP was determined with hexokinase (ATP:D-hexose 6-phosphotransferase, EC 2.7.I.I) and glucose 6-phosphate dehydrogenase (D-glucose 6-phosphate:NADP+ oxidoreductase, EC 1.I.I.49), ADP with pyruvate kinase (ATP:pyruvate phosphotransferase, EC 2.7.I.40) and lactate dehydrogenase (L-lactate:NAD+ oxidoreductase, EC 1.I.I.27), and glutamate and ammonia with glutamate dehydrogenase (L-glutamate:NAD+ oxidoreductase (deaminating), EC 1.4.I.2). All these analyses were carried out as described by Bergmeyer³², except that in the ammonia determination Tris buffer was replaced by 0.3 M potassium phosphate buffer (pH 7.5). Glutamine was determined by adding glutaminase (L-glutamine amidohydrolase, EC 3.5.I.2) after completion of the ammonia determination.

Magnesium was determined by titration with EDTA in ammonia—NH $_4$ Cl buffer (pH 10) with Erio Black T as indicator.

The pH was determined with a Radiometer Titrator Type TTT 1c. The electrode was standardized at pH values of 4.0 and 7.0 with buffersolution tablets supplied

by Burroughs Welcome and Co., London. The temperature was maintained at 25 ± 0.3 °C.

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REFERENCES

- 1 L. Levintow and A. Meister, J. Biol. Chem., 209 (1954) 265.
- 2 T. Benzinger, C. Kitzinger, R. Hems and K. Burton, Biochem. J., 71 (1959) 400.
- 3 R. A. Alberty, J. Biol. Chem., 243 (1968) 1337.
- R. A. Alberty, J. Biol. Chem., 244 (1969) 3290.
 R. C. Phillips, P. George and R. J. Rutman, J. Biol. Chem., 244 (1969) 3330.
- 6 R. S. Cockrell, E. J. Harris and B. C. Pressman, Biochemistry, 5 (1966) 2326.
- 7 E. C. Slater, in L. Ernster and Z. Drahota, Mitochondria-Structure and Function, Academic Press, London and New York, 1969, p. 205.
- 8 E. C. Slater, in S. Papa, J. M. Tager, E. Quagliariello and E. C. Slater, The Energy Level and Metabolic Control in Mitochondria, Adriatica Editrice, Bari, 1969, p. 255.
- 9 E. C. Slater, Q. Rev. Biophys., 4 (1971) 35.
 10 L. J. M. Eilermann and E. C. Slater, Biochim. Biophys. Acta, 216 (1970) 226.
- 11 R. Kraayenhof, Biochim. Biophys. Acta, 180 (1969) 213.
- 12 J. E. Varner and G. C. Webster, Plant Physiol., 30 (1955) 393.
- 13 R. F. Lumb and A. E. Martell, J. Phys. Chem., 57 (1953) 690.
- 14 J. Schubert, J. Phys. Chem., 56 (1952) 113.
- 15 R. A. Alberty, R. M. Smith and R. M. Bock, J. Biol. Chem., 193 (1951) 425.
- 16 R. C. Phillips, P. George and R. J. Rutman, J. Am. Chem. Soc., 88 (1966) 2631.
- 17 C. Kitzinger and R. Hems, Biochem. J., 71 (1959) 395.
- 18 C. Kitzinger and T. Benzinger, Z. Naturforsch., 10 (1955) 375.
- 19 R. J. Podolsky and M. F. Morales, J. Biol. Chem., 218 (1956) 945.
 20 R. C. Phillips, P. George and R. J. Rutman, Biochemistry, 2 (1963) 501.
- 21 J. M. Stokes, Trans. Faraday Soc., 41 (1945) 685.
- 22 G. Scatchard and P. C. Breckenridge, J. Phys. Chem., 58 (1954) 596.
- 23 E. A. Robbins and P. D. Boyer, J. Biol. Chem., 224 (1957) 121.
- 24 M. R. Atkinson, E. Johnson and R. K. Morton, Nature, 184 (1959) 1925.
- 25 J. H. Ashby, H. B. Clarke, E. M. Crook and S. P. Datta, Biochem. J., 59 (1955) 203.
- 26 H. B. Clarke, D. C. Cusworth and S. P. Datta, Biochem. J., 58 (1954) 146.
- 27 R. M. Smith and R. A. Alberty, J. Am. Chem. Soc., 78 (1956) 2376.
- 28 R. M. Smith and R. A. Alberty, J. Phys. Chem., 60 (1956) 180.
- 29 N. C. Melchior, J. Biol. Chem., 208 (1954) 615.
- 30 R. A. Ronzio, W. B. Rowe, S. Wilk and A. Meister, Biochemistry, 8 (1969) 2670.
- 31 J. B. Sumner, Science, 100 (1944) 413.
- 32 H. U. Bergmeyer, Methoden der Enzymatischen Analyse, Band I und Band II, Verlag Chemie, Weinheim/Bergstr., 1970.

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